

Medical Progress

Current Status of Renal Transplantation

MICHAEL G. SURANYI, MBBS, FRACP, and BRUCE M. HALL, MBBS, FRACP, PhD, *Stanford, California*

The success rate of renal transplantation has improved considerably during the past decade, with substantial improvements in both graft and patient survival. The quality of graft function, however, and not graft survival alone is increasingly determining the standards by which transplantation outcome is being judged. As the demand for kidney transplants continues to rise and transplants are being offered to an ever-increasing number of patients, organs are being sought from new supply pools and efforts are being made to use current resources more efficiently. Improvements in clinical management have allowed short-term complications such as infection and rejection to be better prevented or better diagnosed and treated. Fundamental advances in the understanding of the immunologic processes underlying both allograft rejection and acceptance and the introduction of new immunosuppressive agents have allowed a better use of drug therapy and have moved the goal of acquired transplant tolerance closer to attainment. With improved initial transplant success rates, the long-term transplantation outcome is becoming more important. The role of tissue matching in preventing chronic rejection is becoming more appreciated, and the long-term risks of malignancy, arteriosclerosis, and chronic rejection are being better recognized and managed.

(Suranyi MG, Hall BM: Current status of renal transplantation. *West J Med* 1990 Jun; 152:687-696)

In the 1980s the success rate of cadaver-donor renal transplantation improved from one-year graft survival rates of around 50% to rates of 75% to 85%. At the same time, patient survival rates also improved, with one-year patient survival generally greater than 95%.¹⁻³ During the past decade, there have been parallel improvements in the detailed understanding of the fundamental processes causing rejection.^{4,5} In clinical transplantation, improvements have been made in tissue typing, immunosuppression, and patient care and in the management of rejection, the diagnosis and treatment of acute and chronic allograft dysfunction, and the diagnosis and treatment of immunosuppression-related problems such as malignancy and infection.

Despite the continued relative shortage of cadaveric donor kidneys, renal transplantation grafting rates are progressively rising. The number of kidney transplants is increasing as groups of patients previously denied them—including those sensitized to HLA antigens,⁶ the elderly,⁷⁻⁹ the very young,¹⁰ and high-risk patients¹¹—receive kidneys.

The solution to the shortage of kidneys lies first with the optimal use of currently available cadaver kidneys and, second, in harvesting grafts from new patient pools, such as kidneys from pediatric and elderly donors¹²⁻¹⁴ and from living-related and unrelated donors.¹⁵⁻¹⁷

Pretransplantation Measures to Improve Graft Survival

Tissue Typing

The most important transplantation antigens in humans and animal models are determined by a polymorphic group of genes, the major histocompatibility complex (MHC),

which in humans are called human leukocyte antigens. The protein products of these genes are present on most cell surfaces and fall into two important classes: the HLA class I antigens (including HLA-A, -B, and -C) and the HLA class II antigens (including the HLA-DR, -DP, -DQ, and other D-related antigens). A set, or haplotype, of these antigens is inherited from each parent and is codominantly expressed. The resolution of the x-ray crystallographic structure of HLA-A2 has shown that the normal function of HLA molecules is to present antigen, in the form of a short peptide (5 to 22 amino acids), in the groove formed on the external surface of the molecules.¹⁸ T-cell receptors recognize only antigen presented by this complex of MHC and antigenic peptide, and specialized antigen-presenting cells provide activation signals to T cells. This antigen presentation system is designed to trigger the immune system against viral and other intracellular pathogens and tumors. In transplantation, a situation unforeseen by nature, these same mechanisms result in the activation of effector cells such as T and B cells against the foreign MHC on the cells of the allograft, and the end result is rejection.

The stimulus activating the responding immune system to induce rejection, the major cause of allograft loss, depends on the number and type of HLA mismatches between recipient and donor. The most important HLA antigen mismatches driving the rejection response have been found to be the HLA-DR and HLA-B antigens, which are also the most polymorphic.¹⁹ The benefits of HLA matching are best manifested in grafts between relatives, where matching between donor and recipient influences both short-term and long-term allograft survival.¹⁶

ABBREVIATIONS USED IN TEXT

ALG = antilymphocyte globulin
 CMV = cytomegalovirus
 HIV = human immunodeficiency virus
 IL-2 = interleukin 2
 MHC = major histocompatibility complex

With the use of potent immunosuppressive protocols, early graft losses due to rejection can be suppressed, allowing the successful short-term function of even completely mismatched kidneys.^{17,20} With longer follow-up, however, HLA disparity may contribute substantially to the loss of grafts with chronic rejection despite immunosuppression.^{21,22} The rejection of poorly matched grafts results in a high incidence of sensitization, which can jeopardize the chances and success of subsequent transplantation and can result in a high mortality rate among patients returning to dialysis.²³ The success of second transplants may depend even more on HLA matching than does that of primary grafts.²⁴

The need to establish the optimal use of a limited donor resource has led to a promotion of kidney sharing on a regional basis, with the aim of increasing the number of patients receiving well-matched grafts.²⁵⁻²⁷ The national United Network of Organ Sharing program was set up with that aim. By current estimates an effective organ-sharing scheme could result in as many as 20% of patients being allocated A-, B-, and DR-compatible kidneys and in the number of patients receiving poorly matched kidneys being substantially reduced.²⁸ Organ-sharing programs may also have the benefit of reducing the number of highly sensitized patients awaiting second transplants. Nevertheless, kidney sharing is a contentious issue.^{29,30} Some groups maintain that matching confers no benefit in cadaver or even living-related transplants²⁰ and that the longer storage required for exchanged kidneys causes damage that counterbalances any advantages of tissue-matched exchanged kidneys.³¹ There has been progressive improvement, however, in the cold storage preservation of cadaveric kidneys, particularly with the introduction of the University of Wisconsin solution.³²⁻³⁵ Still, it may require the application of exchange incentives to improve cooperation to produce an effective allograft exchange program.³⁶

Lymphocytotoxic Antibody Crossmatch

After excluding ABO blood group incompatibility, testing for lymphocytotoxic antibodies against HLA class I and class II constitutes one of the most important matching procedures before transplantation. Such antibodies arise as a consequence of blood transfusion, pregnancy, and a previous transplant rejection.³⁷ The presence of antibody reacting with donor lymphocytes is excluded before transplantation, as such antibody has been associated with hyperacute and accelerated rejection.³⁸ Transplantation generally does not take place if deleterious lymphocytotoxic antibodies are detected in the crossmatch test before the procedure.³⁹

Numerous controversies remain unresolved regarding the techniques and criteria of such testing,⁴⁰ including the relative value of historic and current antidonor crossmatches, the merits of anti-T- and anti-B-cell crossmatches,⁴¹ the relative importance of sensitive assays such as fluorescence-activated cell sorting,⁴²⁻⁴⁴ and the use of antiglobulin reagent in the

cytotoxic crossmatch.⁴⁵ The crossmatch test does not identify antibodies against organ-specific antigens that are not expressed on donor lymphocytes.⁴⁶ Such antibodies may be found in the serum or allograft of a recipient⁴⁷ and may also play an important role in rejection processes.⁴⁸

An increasing number of patients awaiting transplantation show a high degree of sensitization, which makes it difficult to find a compatible donor. This trend is exacerbated by the number of patients awaiting second transplants after the rejection of poorly matched first grafts. Many efforts to reduce the incidence of such presensitization have been ineffective,⁴⁹ but grafts bearing maternal HLA antigens may be useful in such sensitized recipients.⁵⁰ Highly sensitized patients do best when receiving transplants using well-matched grafts,²⁴ but such patients have also had successful transplants without regard to HLA matching.⁵¹ Attempts have been made to reverse the sensitized state by plasmapheresis, antibody adsorption, and cyclophosphamide therapy, but these are still experimental.^{52,53} Similar protocols have been used to remove hemagglutinins in recipients of ABO-incompatible grafts.

Blood Transfusion

In 1973 Opelz and co-workers noted a beneficial effect of random blood transfusion on the outcome of transplantation,⁵⁴ and this was subsequently confirmed by many groups. The recognition of the therapeutic benefits of random transfusion was followed shortly by the introduction of donor-specific blood transfusions in a subset of living-related transplant recipients.⁵⁵ Both random and donor-specific blood transfusions have been associated with a significant incidence of sensitization,⁵⁶ which can preclude subsequent transplantation. It has been postulated that donor-specific transfusion results in directly sensitizing recipients, whose reactive immune cells are then depleted by heavy immunosuppression soon after the transplantation.⁵⁷

The role of blood transfusion as an adjunct to transplantation is now in question,⁵⁸ and the risk of sensitization and of the transmission of viral and other pathogens—particularly the human immunodeficiency virus (HIV)—must be weighed against the potential benefits. Further, with the introduction of recombinant erythropoietin,⁵⁹ the need to transfuse dialysis patients for anemia will be largely eliminated. If transfusion is required, however, using HLA-matched transfusions may reduce sensitization.⁶⁰ Recent data in the International Collaborative Transplant Study have led Opelz to suggest that there is now no compelling reason to undertake either random or donor-specific transfusion.⁶¹

Immunosuppressive Protocols

Immunosuppressive agents commonly used include azathioprine, prednisone, cyclosporine, antilymphocyte globulin (ALG) preparations, anti-CD3 monoclonal antibody (Orthoclone OKT3), ionizing irradiation,⁶² and alkylating agents such as cyclophosphamide.⁶³ Azathioprine and prednisone were the mainstay of therapy in the 1960s and 1970s, and antilymphocyte globulin was added in the late 1960s. Cyclosporine became available in the 1980s and is now central to most protocols.

The introduction of cyclosporine has been associated with a period of progressive improvement in the success of renal transplantation,⁶⁴ which has also been translated into an improvement in quality of life for patients.^{65,66} The mechanisms

of its actions are still being investigated, but it effectively inhibits the release of cytokines, especially interleukin 2 (IL-2), known to be important in the generation of the rejection response.⁶⁷⁻⁶⁹ The main side effect of cyclosporine is, paradoxically, nephrotoxicity.

Cyclosporine was initially used alone or with prednisone, but to minimize side effects, particularly nephrotoxicity that occurs at high dosage, there has been a trend toward combining agents at a lower dosage to achieve additive or synergistic effects.⁷⁰ The rationale of multiple-agent protocols is that each immunosuppressive acts at different sites in the immune system. By using lower doses of each agent, it is hoped that the side effects of higher doses will be avoided while the benefits of each agent are retained. Triple-drug therapy with azathioprine, prednisone, and cyclosporine has become widely used, as has quadruple-drug therapy, with ALG induction followed by triple-drug therapy.⁷⁰ Despite the general tendency to use multiple agents, some transplant centers continue to use one- or two-drug protocols with comparable graft survival rates.^{71,72} The combination of cyclosporine and azathioprine as maintenance therapy can avoid the long-term use of steroids.^{73,74}

The main factor driving the trend toward multiple-drug therapy is the nephrotoxicity of cyclosporine, the use of which can increase the duration of early nonfunction of a transplanted kidney.⁷⁵ To minimize early nonfunction, some protocols include low doses of cyclosporine whereas others avoid the early use of cyclosporine by using ALG, azathioprine, and prednisone followed by cyclosporine only after adequate renal function occurs.⁷⁰

Combination immunosuppression still does not completely prevent rejection episodes or graft loss, especially in second transplants or in patients who do not have good tissue matching.^{3,76} Combined therapy may also be associated with a substantial risk of patient morbidity from opportunistic infection⁷⁷ or a malignant lesion such as lymphoma.⁷⁸ For this reason prophylactic broad-spectrum antibiotics and acyclovir may be used to prevent infection.⁷⁹ Monitoring cyclosporine levels in whole blood, serum, or plasma to adjust the dosage is widely done to minimize the side effects of using cyclosporine.⁸⁰⁻⁸³ Because of nephrotoxicity, cost, and the unknown effects of long-term cyclosporine use, some centers have begun converting from cyclosporine to conventional azathioprine and prednisone therapy.⁸⁴ The main reason for converting from initial cyclosporine therapy is the relatively high incidence of cyclosporine nephrotoxicity and its progressive and ultimately irreversible nature.⁸⁵ The conversion from cyclosporine use can result in improved renal function and blood pressure control,^{71,86,87} and these benefits appear to persist.⁸⁷ There can be a major risk of acute rejection and loss of the graft if conversion is undertaken before rejection is carefully excluded, however.^{70,88,89} Patients undergoing conversion for refractory rejection have a high risk of graft loss.

Although cyclosporine therapy is undoubtedly effective against cell-mediated acute rejection, its effect on chronic rejection is less certain,⁹⁰ and distinguishing chronic rejection from nephrotoxicity may be difficult.

Monoclonal Antibodies

Monoclonal antibody therapy, the most successful example of which is OKT3, became available shortly after cyclosporine was introduced. Polyclonal antilymphocyte antibody preparations (ALG) have had a place in renal transplantation

for many years and continue to contribute to immunosuppression protocols.⁹¹ Preparations of ALG, however, have suffered from variation between batches and from side effects associated with their many specificities.⁷⁰ Now monoclonal antibodies are being developed that have greater purity, specificity, and reproducible biologic activity. Their use has provided a mechanism for precisely targeting antibody-mediated effects to one cell surface molecule. In renal transplantation this allows immunosuppressive therapy that is more specific than using agents such as azathioprine, steroids, and cyclosporine. On the T-cell surface many antigens provide a target for such specific therapy.⁹² Of these, an antibody against CD3 is already in general use, and monoclonal antibodies against the IL-2 receptor and against the T12 molecule have been tested.⁹³ The potential usefulness of the anti-CD4 antibody has not been overlooked,⁹⁴ and efforts are being directed to develop many other monoclonal antibodies for clinical use, including those against activation antigens on T cells.⁹⁵ In addition, monoclonal antibodies against cytokines, such as interleukin 2 and interferon gamma, also require assessment as immunosuppressives in renal transplantation.⁹⁶ Investigation is also underway into the use of monoclonal antibodies, such as those to passenger leukocytes, to reduce the immunogenicity of grafts.⁹⁷

OKT3 Monoclonal Antibody Therapy

OKT3, a murine monoclonal antibody of the immunoglobulin G2a class, has proved to be a potent immunosuppressive agent owing to its ability to bind the T-cell antigen-receptor complex.⁹⁸ The use of OKT3 has been effective as prophylactic therapy in renal transplantation^{99,100} and as anti-rejection therapy,¹⁰¹⁻¹⁰⁵ but its most appropriate use may be as rescue therapy for steroid-resistant rejection.¹⁰⁶⁻¹⁰⁸ OKT3 therapy has also been shown to benefit patients with predominantly vascular pattern rejection.¹⁰⁹ The recurrence rate of rejection, however, may be unexpectedly high after OKT3 use, as it probably does not eliminate allograft-responding T cells. Its use is thought to result initially in the opsonization and rapid removal of T cells from the circulation. Thereafter, OKT3 appears to rapidly modulate or remove the CD3 or T cell-receptor complex from the surface of T cells, rendering them incapable of activation by antigen recognition.

A number of major problems have emerged with the use of OKT3. A severe febrile reaction and bronchospastic response can follow the administration of the first dose. Pulmonary edema also occurs in patients with fluid overload.¹¹⁰ The cause of systemic symptoms may be related to the demonstrated ability of OKT3 to activate T cells. This initial T-cell activation, or, alternatively, a wave of T-cell destruction, can result in the release of potent T-cell cytokines leading to systemic effects, including a fall in the glomerular filtration rate.^{98,111} Cytokine increases have also been noted with ALG use.¹¹² After OKT3 use, particularly if high-dose immunosuppressive agents have also been used, a higher incidence of infection may be seen.^{107,113,114} Finally, under certain circumstances, T cells with modulated CD3 or T cell-receptor complex may still mediate acute rejection, despite therapeutic levels of OKT3.¹¹⁵

An antibody response to OKT3 develops in most treated patients, despite concomitant immunosuppression with cyclosporine or azathioprine. These antibodies may be to the idiotype, isotype, or common determinants of mouse immunoglobulin.⁹⁸ Such antibodies may develop during an ini-

tial course, limiting its usefulness and allowing rejection to break through, but they more often develop afterwards, interfering with the use of a second course.^{116,117} Reuse can be successful if high-titer anti-OKT3 antibodies have not developed,^{118,119} although such reuse is associated with a high incidence of viral infection.¹²⁰

Because anti-idiotypic antibodies developing after OKT3 therapy do not cross-react with all anti-CD3 monoclonal antibodies, using different anti-CD3 monoclonal antibodies may be one approach to reuse.¹²¹ Further, with the use of molecular biologic techniques, human or mouse-human hybrid antibodies can be made that may not stimulate anti-mouse antibody formation.⁹² It has been recently suggested that a concurrent infusion of anti-CD4 monoclonal antibody may reduce the formation of recipient anti-OKT3 antibodies by impairing helper T-cell function required for antibody formation.¹²²

Anti-Interleukin 2-Receptor Therapy

Interleukin 2 is a critical cytokine involved with the activation, proliferative expansion, and amplification of T cell-mediated rejection responses, such as those occurring with allograft rejection.¹²³ The IL-2 receptor is expressed mainly on activated T cells, having a low- and high-affinity form, the latter of which binds and internalizes IL-2. In patients undergoing transplantation, the blockade or elimination of T cells expressing the IL-2 receptor can be expected to immunosuppress, in a relatively selective way, the clones responding to an allograft and mediating rejection.¹²⁴ The use of anti-IL-2 receptor monoclonal antibodies has proved effective in animal models¹²⁵ and has undergone initial trials in humans,^{126,127} but not always successfully.¹²⁸ This therapy may be synergistic with cyclosporine therapy and is reported to spare suppressor cells.¹²⁹

The initial promise of monoclonal antibody therapy using anti-IL-2 receptors has been enlarged on by the development of a novel IL-2-diphtheria toxin hybrid, which binds to the high-affinity IL-2 receptor on activated cells, mainly T cells, is internalized, and results in cell destruction.¹³⁰⁻¹³²

Newer Agents

The promise of less nephrotoxic analogues of cyclosporine has not yet been realized.¹³³ With immunosuppressive characteristics similar to cyclosporine, FK 506 has recently come to increased public attention. Similar to cyclosporine in its ability to inhibit cytokine production,¹³⁴⁻¹³⁶ FK 506 is currently under investigation for use as a clinical renal transplant immunosuppressive agent, but it has shown considerable hepatotoxicity.¹³⁷ Investigation is also proceeding into methods of treating allografts before implantation with agents designed to reduce the ability of grafts to stimulate an immune response.^{138,139}

Efforts to Induce Transplant Tolerance

The development of tolerance, or specific unresponsiveness, to donor tissue is the most attractive form of immunosuppression, avoiding drug complications such as infection and malignancy. Anecdotes relate long-term survival of some allografts without the benefit of continued immunosuppressive therapy for prolonged periods.¹⁴⁰ With the establishment of tolerance, immunosuppressive drugs would not be required, and the immune system's natural regulatory mechanism would ensure allograft survival. This ideal is still far

from reality in clinical transplantation, although our increased understanding of the mechanisms of such states in animals brings it closer.¹⁴¹

In experiments in animals, total lymphoid irradiation and donor bone marrow infusion have been shown to promote a tolerant state, and the role of these agents is being investigated in humans.^{142,143} Recent reviews of total lymphoid irradiation in a clinical transplant population found some patients with unresponsiveness to the donor in *in vitro* immune responses, suggesting that a state of specific donor unresponsiveness may develop in some patients.^{144,145} Total lymphoid irradiation is effective immunosuppression, but long-term follow-up shows that patients receiving this therapy have three-year patient and graft survivals, which appear lower than the results of comparable trials using cyclosporine and prednisone.¹⁴⁶

Clinical Problems in Renal Transplantation

The Diagnosis of Rejection

The diagnosis and treatment of renal dysfunction early in the posttransplant period are difficult but important, because acute tubular necrosis, cyclosporine nephrotoxicity, and acute rejection are hard to distinguish clinically.¹⁴⁷ Percutaneous renal biopsy remains the gold standard for diagnosing the cause of renal transplant impairment, especially with the addition of monoclonal antibody markers to identify infiltrating cells and allograft antigens.¹⁴⁸⁻¹⁵¹ In particular, the assessment of HLA-DR expression on renal tubular cells may assist in the determination of acute rejection.¹⁵²

Fine-needle aspiration biopsy has become a clinically useful and safe technique for sequentially monitoring graft parenchymal cells and infiltrates.¹⁵³ The application of flow cytometric and immunofluorescence technology has added increased usefulness to the technique^{154,155}; although the correlation with renal allograft histologic features is good, the information provided about graft structures is more limited.

Renal ultrasonography, radionuclide studies, and magnetic resonance imaging (MRI) may all contribute to the diagnosis of renal transplant dysfunction. Nevertheless, despite their technologic sophistication, none of these methods are sufficient to distinguish among rejection, acute tubular necrosis, and cyclosporine nephrotoxicity. Rejection causes a loss of the corticomedullary demarcation on MRI, but this is not specific.¹⁵⁶⁻¹⁵⁸ Whether MRI is more sensitive than radionuclide studies and sonography has been disputed.¹⁵⁹⁻¹⁶¹

Techniques requiring further assessment include duplex Doppler sonography, which may aid in differentiating acute rejection from acute tubular necrosis or cyclosporine nephrotoxicity.¹⁶²⁻¹⁶⁴

Cyclosporine Nephrotoxicity

Cyclosporine nephrotoxicity remains a major problem in renal transplantation, and efforts to resolve whether the immunosuppressive effect of cyclosporine can be separated from its nephrotoxic effect have so far failed.¹⁶⁵ The therapeutic window is so narrow that cyclosporine nephrotoxicity and allograft rejection may coexist. Cyclosporine dosage and cyclosporine serum concentrations may not provide accurate predictors of nephrotoxicity¹⁶⁶⁻¹⁶⁸; nevertheless, many groups advocate protocols using lower cyclosporine dosages to lessen renal impairment.^{169,170}

Cyclosporine can induce nephrotoxicity in both native

and transplanted kidneys,¹⁷¹ but nephrotoxicity may be worse in kidneys that have had an ischemic insult.¹⁷² Patients receiving cyclosporine have a higher incidence and longer duration of initial graft nonfunction,^{71,173} which may result in long-term effects on graft survival.¹⁷⁴ For this reason some centers prefer to use sequential therapy, treating initially with other immunosuppressive agents and introducing a regimen of cyclosporine after graft function has been established.¹⁷⁵ Further, patients with functioning grafts who are receiving long-term cyclosporine therapy generally have a lower glomerular filtration rate than do patients receiving azathioprine and prednisone therapy.⁷¹

The exact mechanisms of cyclosporine nephrotoxicity are still not known but are under intensive investigation. Cyclosporine causes vasoconstriction of vascular smooth muscle,¹⁷⁶ with an alteration in renal hemodynamics,¹⁷⁷ whereas tubular function is relatively spared.¹⁷⁸ In addition, cyclosporine may be responsible for the arteriopathy found in some allografts.¹⁷⁹ Cyclosporine may alter the thromboxane-prostacyclin balance, allowing thromboxane-induced vasoconstrictor predominance.¹⁸⁰⁻¹⁸³ Certainly cyclosporine use appears to increase thromboxane A₂ levels,¹⁸⁴ and administering thromboxane inhibitors modifies the renal effects of cyclosporine.^{185,186} Because cyclosporine is metabolized in the liver through the cytochrome P-450 system, drug interactions may be important in inducing elevated cyclosporine levels predisposing to nephrotoxicity. The use of erythromycin, ketoconazole, steroids, sex hormones, and some calcium channel blockers may induce elevated cyclosporine levels, and rifampin and anticonvulsant therapy may decrease cyclosporine concentrations.¹⁸⁷

The use of calcium channel blockers may modify cyclosporine nephrotoxicity¹⁸⁸ because cyclosporine binds certain intracellular calcium-binding proteins.¹⁸⁹ Although it is clear that cyclosporine therapy results in lower kidney transplant function than does azathioprine and prednisone therapy,⁷¹ its role in progressive renal impairment is in question.¹⁹⁰ It is best documented that long-term cyclosporine administration may be associated with progressive renal impairment in patients receiving organs other than kidneys,^{191,192} where renal transplant rejection does not confuse the issue. In the long term, cyclosporine use is associated with a progressive increase in interstitial fibrosis and tubular and vascular changes.¹⁹³ Even when creatinine levels are stable, histologic damage can be detected in the allograft, and stable creatinine levels may be at the cost of increased function in hypertrophied remnant glomeruli.^{85,191,194} Thus, the use of serum creatinine levels or even creatinine clearance as an indicator of kidney transplant function may not be sensitive to progressing nephrotoxicity.¹⁹⁵

Transplant-Associated Hypertension

Hypertension has been a recurrent and persistent problem in renal transplantation.¹⁹⁶ The incidence of posttransplantation hypertension has increased since the widespread use of cyclosporine.^{197,198} In association with the impaired lipid profile related to cyclosporine use (reviewed by Chapman and Morris),⁸⁴ hypertension may contribute substantially to the death of transplant recipients through the development of arteriovascular and cardiovascular disease.^{199,200} Hypertension may also pose a threat to allograft survival.^{23,201} The basis of the high incidence of hypertension in renal transplant patients includes factors such as chronic rejection, transplant

renal artery stenosis, recurrent renal disease, and increased native-kidney renin production.^{202,203} Cyclosporine is a potent inducer of hypertension, in a somewhat different manner from other immunosuppressive agents.^{198,204,205} Cyclosporine causes subtle renal impairment,²⁰⁶ even at normal therapeutic levels,²⁰⁷ and its use has been associated with sodium retention.²⁰⁸ The addition of prednisone therapy can also exacerbate hypertension, in part by limiting renal sodium excretion.²⁰⁹

Opportunistic Infection

Viral infections have the greatest potential for causing serious infectious complications in kidney transplant recipients. There is also increasing recognition of the role of viruses in triggering posttransplant malignancy, such as lymphoma, hepatoma, Kaposi's sarcoma, and genitourinary cancers.^{210,211} Viruses in the herpesvirus group include herpes simplex, herpes zoster, Epstein-Barr virus, and cytomegalovirus (CMV). Cytomegalovirus remains one of the major infectious complications in renal transplantation, where the source of the virus may be the donor organ, blood transfusion, or reactivation of latent virus in the recipient.²¹² Cytomegalovirus infections may be deleterious to graft survival and function and may cause substantial patient morbidity and mortality.²¹³ In most cases primary infections are more severe than reactivation of a latent virus, and both antilymphocyte globulin and OKT3 therapy can potentiate CMV infections.^{113,214} It is now recognized that reinfection with a second virus strain may occur in seropositive recipients and that such patients are thus not immune to repeated primary CMV infections.²¹⁵ Reserving CMV-negative donors for CMV-negative recipients appears to be a useful preventive measure where practical but may add difficulty to transplant matching.²¹⁶⁻²¹⁸ Efforts to develop an effective vaccine against CMV have been mostly disappointing,²¹⁹⁻²²¹ but passive immunization with CMV hyperimmune globulin seems effective.²²² Prophylactic therapy with oral acyclovir after transplantation has reduced the incidence of primary CMV infection, and its use may reduce that of Epstein-Barr virus and herpesvirus infection as well as the incidence of Epstein-Barr-related lymphoproliferative disorders.²²³ Because rapid diagnosis is critical, new and quicker tests to diagnose CMV infection are being introduced.²²⁴⁻²²⁶ In addition, anti-CMV therapy has recently improved.²²⁷ Whereas CMV lacks the virus-specific thymidine kinase needed for the phosphorylation and activation of acyclovir, newer agents such as ganciclovir are particularly active against CMV.²²⁸⁻²³⁰ Other antiviral agents such as foscarnet²³¹ are also proving effective. There may be a role for combining therapy with hyperimmune globulin and ganciclovir in high-risk patients with serious infections.²³²

The herpes simplex virus can also be transmitted by the renal allograft,²³³ but oral acyclovir has been used with good results to prevent herpes simplex infection in kidney transplant recipients.²³⁴

Transmission of the human immunodeficiency virus with the transplant organ or blood transfusion can occur despite the screening of donors.²³⁵⁻²³⁷ Diagnosing the acquired immunodeficiency syndrome in persons with transplants may be more difficult, as antibody responses to HIV may be impaired.²³⁸ Patients infected with HIV have a poor prognosis.^{239,240} No patient with HIV antibodies should receive or donate an organ.²³⁵ Patients with seroconversion after trans-

plantation should have their immunosuppression reduced, and possibly stopped, but they almost inevitably do poorly.²⁴¹

Malignancy

The increased incidence of malignancy in patients receiving kidney transplants has been well documented^{210,242} and is due to immunosuppressive therapy, viral infections, and occasionally to the transmission of cancer with the donor organs.²⁴³ With increasing potency of immunosuppressive medication, the incidence of malignancy in transplant patients has increased.²⁴⁴

Skin cancer is the most common manifestation in some countries.^{245,246} Epstein-Barr virus-related lymphoma has been of recent concern, and its appearance is associated with potent immunosuppression, especially with multiple drugs and high doses.^{78,247,248} Some of these polyclonal lymphoproliferative states are responsive to acyclovir therapy combined with reduced immunosuppression.²⁴⁹

Long-term Outcome

The major causes of graft loss in the long term are chronic rejection and patient death. The predominant causes of death are infections, malignant neoplasms, and arteriovascular disease.^{210,250,251}

When rejection takes place in long-surviving grafts, it is generally chronic or vascular pattern rejection, although acute rejection can also occur late.²⁵² Chronic rejection is poorly understood. It generally presents as a slow deterioration in renal function and less often as a sequence of episodes of renal impairment, but both respond poorly to antirejection therapy. Grafts with chronic rejection show progressive pruning of the distal arterial tree. A biopsy specimen shows a vascular pattern of rejection, with fibroblastic endothelial thickening of interlobular and arcuate arteries as well as glomerular and tubular ischemic changes.²⁵³ While conventional theory holds that chronic rejection is predominantly antibody-mediated, evidence is growing that cellular immunity may play an important role.²⁵⁴

Chronic rejection is responsible for a significant proportion of late graft losses and contributes to a 3% to 10% annual late graft failure rate.^{209,255} Among these losses must be counted grafts rejected because of poor patient compliance,^{256,257} which may occur particularly in children and adolescents,²⁵⁸ sometimes because of changes in appearance induced by therapy.^{259,260} Glomerulopathy in kidney transplants may represent rejection, infection,²¹⁴ recurrent primary disease, or de novo glomerulonephritis. Some forms of glomerulonephritis, such as focal sclerosing glomerulonephritis,²⁶¹ IgA disease,²⁶² and membranoproliferative glomerulonephritis (type II), have a tendency to recur, often with deleterious consequences, and a proportion of patients go on to lose their grafts.^{210,263,264} Immunosuppressive protocols including cyclosporine appear not to be effective in preventing the recurrence of glomerulonephritis in transplanted kidneys.²⁶⁵

Future Directions

In the past decade there have been remarkable improvements in the understanding of the mechanisms of rejection and the actions of immunosuppressive agents. In conjunction with better management of infection and better patient care, these factors have been translated into better graft and patient survival after renal transplantation. It is now important to

focus on the issues of patient well-being and the level of kidney function.

In the next decade further important advances in renal transplantation should occur, with an array of new immunosuppressive agents, better diagnostic abilities, and better antibiotics. The ability to induce specific transplant tolerance, with a reduction or elimination of the need for nonspecific immunosuppressive agents, has been shown in animal models and needs to be adapted to humans, as suggested by Strober and colleagues.¹⁴⁵ The ability to harness the immune system's own immunoregulatory processes to produce an acquired specific unresponsiveness to organ allografts could well eliminate many of the problems limiting the success of renal transplantation.

REFERENCES

1. Kootte AM, Lensen LM, van Bockel JH, et al: High- and low-dose regimens of cyclosporin in renal transplantation: Immunosuppressive efficacy and side effects. *Nephrol Dial Transplant* 1988; 3:666-670
2. Najarian JS, Canafax DM, Sutherland DE: Renal transplantation in diabetic patients is confirmed therapy while pancreas transplantation should be performed only in an investigational setting. *J Diabetic Complications* 1988; 2:158-161
3. Ponticelli C, Tarantino A, Montagnino G, et al: A randomized trial comparing triple-drug and double-drug therapy in renal transplantation. *Transplantation* 1988; 45:913-918
4. Krensky AM, Suranyi MG, Clayberger C, et al: Advances in the immunobiology and clinical practice of transplantation. *Curr Nephrol* 1989; 13:417-465
5. Halloran PF, Cockfield SM, Madrenas J: The molecular immunology of transplantation and graft rejection. *Immunol Allergy Clin North Am* 1989; 9:1-20
6. Claas FH, Van Rood JJ: Transplantation in hyperimmunized patients. *Adv Nephrol* 1989; 18:317-323
7. Pirsch JD, Stratta RJ, Armbrust MJ, et al: Cadaveric renal transplantation with cyclosporine in patients more than 60 years of age. *Transplantation* 1989; 47:259-261
8. Shah B, First MR, Munda R, et al: Current experience with renal transplantation in older patients. *Am J Kidney Dis* 1988; 12:516-523
9. Lauffer G, Murie JA, Gray D, et al: Renal transplantation in patients over 55 years old. *Br J Surg* 1988; 75:984-987
10. Clark AGB, Rigden SPA, Haycock GB, et al: Renal transplantation in children. *Transplant Rev (Orlando)* 1987; 1:101-131
11. Kaufman DB, Sutherland DER, Fryd DS, et al: A single-center experience of renal transplantation in thirteen Jehovah's Witnesses. *Transplantation* 1988; 45:1045-1049
12. Smith AY, Van Buren CT, Lewis RM, et al: Short-term and long-term function of cadaveric kidneys from pediatric donors in recipients treated with cyclosporine. *Transplantation* 1988; 45:360-367
13. Fine RN: In depth review: Renal transplantation of the infant and young child and the use of pediatric cadaver kidneys for transplantation in pediatric and adult recipients. *Am J Kidney Dis* 1988; 12:1-10
14. Kasiske BL: The influence of donor age on renal function in transplant recipients. *Am J Kidney Dis* 1988; 11:248-253
15. Donnelly PK, Clayton DG, Simpson AR: Transplants from living donors in the United Kingdom and Ireland: A centre survey. *Br Med J [Clin Res]* 1989; 298:490-493
16. Tilney NL, Hollenberg NK: Use of living donors in renal transplantation. *Transplant Rev (Orlando)* 1987; 1:225-238
17. Pirsch JD, Sollinger HW, Kalayoglu M, et al: Living-unrelated renal transplantation: Results in 40 patients. *Am J Kidney Dis* 1988; 12:499-503
18. Bjorkman PJ, Saper MA, Samraoui B, et al: The foreign antigen binding site and T-cell recognition regions of class I histocompatibility antigens. *Nature* 1987; 329:512-523
19. Albert ED: What is new in HLA in 1988? *Transplant Rev (Orlando)* 1988; 2:207-219
20. Kaufman DB, Sutherland DER, Noreen H, et al: Renal transplantation between living-related sibling pairs matched for zero-HLA haplotypes. *Transplantation* 1989; 47:113-119
21. Lewis RM, Janney RP, Golden DL, et al: Stability of renal allograft function associated with long-term cyclosporine immunosuppressive therapy—Five-year follow-up. *Transplantation* 1989; 47:266-272
22. Takiff H, Cook D, Himaya N, et al: Dominant effect of histocompatibility on ten-year kidney transplant survival. *Transplantation* 1988; 45:410-415
23. Wynn JJ, Pfaff WW, Patton PR, et al: Late results of renal transplantation. *Transplantation* 1988; 45:329-333
24. Opelz G: Influence of HLA matching on survival of second kidney transplants in cyclosporine-treated recipients. *Transplantation* 1989; 47:823-827
25. Gilks WR, Bradley BA, Gore SM, et al: Substantial benefits of tissue matching in renal transplantation. *Transplantation* 1987; 43:669-674
26. Opelz G: The benefit of exchanging donor kidneys among transplant centers. *N Engl J Med* 1988; 318:1289-1292
27. Takiff H, Iwaki Y, Cecka M, et al: The benefit and underutilization of sharing kidneys for better histocompatibility. *Transplantation* 1989; 47:102-105
28. Cicciarelli J, Terasaki PI, Mickey MR: The effect of zero HLA class I and II mismatching in cyclosporine-treated kidney transplant patients. *Transplantation* 1987; 43:636-640

29. Hunsicker LG: Renal transplantation for the nephrologist: HLA matching and cadaveric kidney allocation (Editorial). *Am J Kidney Dis* 1989; 13:438-441
30. Salvatierra O: Optimal use of organs for transplantation. *N Engl J Med* 1988; 318:1329-1331
31. Alexander JW, Vaughn WK, Pfaff WW: Local use of kidneys with poor HLA matches is as good as shared use with good matches in the cyclosporine era: An analysis at one and two years. *Transplant Proc* 1987; 19:672-674
32. Hoffmann RM, Stratta RJ, D'Alessandro AM, et al: Combined cold storage-perfusion preservation with a new synthetic perfusate. *Transplantation* 1989; 47:32-37
33. McNulty JF, Ploeg RJ, Southard JH, et al: Successful five-day perfusion preservation of the canine kidney. *Transplantation* 1989; 47:37-41
34. Henry ML, Sommer BG, Ferguson RM: Improved immediate function of renal allografts with Belzer perfusate. *Transplantation* 1988; 45:73-75
35. Belzer FO, Southard JH: Principles of solid-organ preservation by cold storage. *Transplantation* 1988; 45:673-676
36. O'Brien BJ: A game-theoretic approach to donor kidney sharing. *Soc Sci Med* 1988; 26:1109-1116
37. Burlingham WJ, Stratta R, Mason B, et al: Risk factors for sensitization by blood transfusions. *Transplantation* 1989; 47:140-144
38. Scornik JC, Salomon DR, Lim PB, et al: Posttransplant antidonor antibodies and graft rejection—Evaluation by two-color flow cytometry. *Transplantation* 1989; 47:287-290
39. Baldwin WM, Paul LC, Claas FH, et al: Destructive and protective effects of antibody on transplants in humans: Practical and theoretical considerations. *Prog Clin Biol Res* 1986; 224:41-57
40. Carpenter CB, Milford EL: HLA matching in cadaveric renal transplantation. *Immunol Allergy Clin North Am* 1989; 9:45-60
41. Taylor CJ, Chapman JR, Fuggle SV, et al: A positive B-cell crossmatch due to IgG anti-HLA-DQ antibody present at the time of transplantation in a successful renal allograft. *Tissue Antigens* 1987; 30:104-112
42. Talbot D, Givan AL, Shenton BK, et al: The relevance of a more sensitive crossmatch assay to renal transplantation. *Transplantation* 1989; 47:552-555
43. Talbot D, Givan AL, Shenton BK, et al: Rapid detection of low levels of donor specific IgG by flow cytometry with single and dual colour fluorescence in renal transplantation. *J Immunol Methods* 1988; 112:279-283
44. Lazda VA, Pollak R, Mozes MF, et al: The relationship between flow cytometer crossmatch results and subsequent rejection episodes in cadaver renal allograft recipients. *Transplantation* 1988; 45:562-565
45. Stratta RJ, Mason B, Lorentzen DF, et al: Cadaveric renal transplantation with quadruple immunosuppression in patients with a positive antiglobulin crossmatch. *Transplantation* 1989; 47:282-286
46. Joyce S, Flye MW, Mohanakumar T: Characterization of kidney cell-specific, non-major histocompatibility complex alloantigen using antibodies eluted from rejected human renal allografts. *Transplantation* 1988; 46:362-369
47. Evans PR, Trickett LP, Gosney AR, et al: Detection of kidney-reactive antibodies at crossmatch in renal transplant recipients. *Transplantation* 1988; 46:844-852
48. Cerilli J, Clarke J, Doolin T, et al: The significance of a donor-specific vessel crossmatch in renal transplantation. *Transplantation* 1988; 46:359-361
49. Weir MR, Shen SY, Dagher FJ, et al: Brief communications: Effects of allostimulation and cyclosporine therapy on cytotoxic antibody production in highly sensitized prospective renal transplant recipients. *Transplantation* 1988; 46:591-593
50. Claas FH, Gijbels Y, van Der Velden-de Munck J, et al: Induction of B cell unresponsiveness to noninherited maternal HLA antigens during fetal life. *Science* 1988; 241:1815-1817
51. Matas AJ, Tellis VA, Quinn TA, et al: Successful transplantation of highly sensitized patients without regard to HLA matching. *Transplantation* 1988; 45:338-342
52. Taube D, Palmer A, Welsh K, et al: Removal of anti-HLA antibodies prior to transplantation: An effective and successful strategy of highly sensitized renal allograft recipients. *Transplant Proc* 1989; 21:694-695
53. Palmer A, Taube D, Welsh K, et al: Removal of anti-HLA antibodies by extracorporeal immunoadsorption to enable transplantation. *Lancet* 1989; 1:10-12
54. Opelz G, Sengar PD, Mickey MR, et al: Effect of blood transfusions on subsequent kidney transplants. *Transplant Proc* 1973; 5:253-259
55. Cochrum KC, Hanes D, Potter D, et al: Donor-specific blood transfusions in HLA-D disparate one-haplotype related allografts. *Transplant Proc* 1979; 11:1903-1907
56. Pfaff WW, Howard RJ, Scornik JC, et al: Incidental and purposeful random donor blood transfusion—Sensitization and transplantation. *Transplantation* 1989; 47:130-133
57. Burlingham WJ, Grailer A, Sordel PM, et al: Improved renal allograft survival following donor-specific transfusions—III. Kinetics of mixed lymphocyte responses before and after transplantation. *Transplantation* 1988; 45:127-132
58. Opelz G: Improved kidney graft survival in nontransfused recipients. *Transplant Proc* 1987; 19:149-152
59. Eschbach JW: Nephrology forum: The anemia of chronic renal failure: Pathophysiology and the effects of recombinant erythropoietin. *Kidney Int* 1989; 35:134-148
60. Scornik JC, Salomon RJ, Pfaff WW: Prevention of transfusion-induced broad sensitization in renal transplant candidates. *Transplantation* 1989; 47:617-620
61. Opelz G: To transfuse or not before transplantation. *Transplant Rev (Orlando)* 1988; 2:77-85
62. Kim TH, Shank BM, Valleria DA, et al: Immunosuppressive techniques using radiation. *Am J Clin Oncol* 1988; 11:362-367
63. Yadav RVS, Indudhara R, Kumar P, et al: Cyclophosphamide in renal transplantation. *Transplantation* 1988; 45:421-424
64. Thorogood J, Van Houwelingen JC, Van Rood JJ, et al: Time trend in annual kidney graft survival. *Transplantation* 1988; 46:686-690
65. Simmons RG, Abress L, Anderson CR: Quality of life after kidney transplantation—A prospective, randomized comparison of cyclosporine and conventional immunosuppressive therapy. *Transplantation* 1988; 45:415-421
66. Rees L, Greene SA, Adlard P, et al: Growth and endocrine function after renal transplantation. *Arch Dis Child* 1988; 63:1326-1332
67. Bishop GA, Hall BM: Effects of immunosuppressive drugs on functions of activated T lymphocytes—Cyclosporine inhibition of gamma interferon production in the presence of interleukin. *Transplantation* 1988; 45:967-972
68. McKenna RM, Szturm K, Jeffery JR, et al: Inhibition of cytokine production by cyclosporine A and G. *Transplantation* 1989; 47:343-348
69. Foxwell BMJ, Ryffel B: The mechanisms of action of cyclosporine. *Immunol Allergy Clin North Am* 1989; 9:79-93
70. Henry ML, Bowers VD, Sommer BG, et al: Combination drug therapies for immunosuppression in renal transplantation. *Transplant Rev (Orlando)* 1988; 2:55-76
71. Hall BM, Tiller DJ, Hardie I, et al: Comparison of three immunosuppressive regimens in cadaver renal transplantation: Long-term cyclosporine, short-term cyclosporine followed by azathioprine and prednisolone, and azathioprine and prednisolone without cyclosporine. *N Engl J Med* 1988; 318:1499-1507
72. Vanderwerf BA, Serota AI: Low-dose cyclosporine for cadaveric renal transplantation. *Transplantation* 1988; 45:320-323
73. Stratta RJ, Armbrust MJ, Oh CS, et al: Withdrawal of steroid immunosuppression in renal transplant recipients. *Transplantation* 1988; 45:323-328
74. Kupin W, Venkat KK, Oh HK, et al: Complete replacement of methylprednisolone by azathioprine in cyclosporine-treated primary cadaveric renal recipients. *Transplantation* 1988; 45:53-55
75. Hall BM, Tiller DJ, Duggin GG, et al: Post-transplant acute renal failure in cadaver renal recipients treated with cyclosporine. *Kidney Int* 1985; 28:178-186
76. D'Alessandro AM, Pirsch JD, Stratta RJ, et al: OKT3 salvage therapy in quadruple immunosuppressive protocol in cadaveric renal transplantation. *Transplantation* 1988; 47:297-300
77. Stratta RJ, Sollinger HW, D'Alessandro AM, et al: Experience with quadruple immunosuppressive therapy in renal transplants. *Immunol Allergy Clin North Am* 1989; 9:109-135
78. Wilkinson AH, Smith JL, Hunsicker LG, et al: Increased frequency of post-transplant lymphomas in patients treated with cyclosporine, azathioprine and prednisone. *Transplantation* 1989; 47:293-296
79. Higgins RM, Bloom SL, Hopkin JM, et al: The risks and benefits of low-dose cotrimoxazole prophylaxis for *Pneumocystis pneumonia* in renal transplantation. *Transplantation* 1989; 47:558-560
80. Keown PA: Optimizing cyclosporine therapy: Dose, levels and monitoring. *Transplant Proc* 1988; 20(suppl 2):382-389
81. Scoutas DS, Hammarstrom M: Comparison of specific radioimmunoassays for cyclosporine. *Transplantation* 1989; 47:668-670
82. Cantarovich F, Bizollon C, Cantarovich D, et al: Cyclosporine plasma levels six hours after oral administration—A useful tool for monitoring therapy. *Transplantation* 1988; 45:389-394
83. Schroeder TJ, Brunson ME, Pesce AJ, et al: A comparison of the clinical utility of the radioimmunoassay, high-performance liquid chromatography, and TDx cyclosporine assays in outpatient renal transplant recipients. *Transplantation* 1989; 47:262-266
84. Chapman JR, Morris PJ: Cyclosporine conversion. *Transplant Rev (Orlando)* 1987; 1:197-224
85. Myers BD, Newton L, Boshkos C, et al: Chronic injury of human renal microvessels with low-dose cyclosporine therapy. *Transplantation* 1988; 46:694-703
86. Hoitsma AJ, Wetzels JFM, van Lier HJJ, et al: Cyclosporin treatment with conversion after three months versus conventional immunosuppression in renal allograft recipients. *Lancet* 1987; 1:584-586
87. Koote AM, Lensen LM, Van Es LA, et al: Controlled cyclosporine conversion at three months after renal transplantation. *Transplantation* 1988; 46:677-680
88. Shen SY, Weir MW, Coughlin TR: Renal allograft biopsy and conversion of cyclosporine to azathioprine. *Transplantation* 1989; 47:223-229
89. Gonwa TA, Nghiem DD, Schulak JA, et al: Results of conversion from cyclosporine to azathioprine in cadaveric renal transplantation. *Transplantation* 1987; 43:225-228
90. Rosenblum ND, Harmon WE, Levey RH: Brief communications: Treatment of chronic renal allograft rejection with cyclosporine and prednisone. *Transplantation* 1988; 45:232-234
91. Matas AJ, Tellis VA, Quinn TA, et al: Individualization of immediate post-transplant immunosuppression—The value of antilymphocyte globulin in patients with delayed graft function. *Transplantation* 1988; 45:406-409
92. Waldmann H: Monoclonal antibodies for organ transplantation: Prospects for the future. *Am J Kidney Dis* 1988; 11:154-158
93. Kirkman RL, Araujo JL, Busch GJ, et al: Treatment of acute renal allograft rejection with monoclonal anti-T12 antibody. *Transplantation* 1983; 36:620-626
94. Hall BM: Therapy with monoclonal antibodies to CD4: Potential not appreciated? *Am J Kidney Dis* 1989; 14(suppl 2):71-77
95. Aversa GG, Suranyi MG, Waugh JA, et al: Detection of late lymphocyte activation marker by A1, a new monoclonal antibody. *Transplant Proc* 1988; 20:49-52
96. Didlake RH, Kim EK, Sheehan K, et al: Brief communications: Effect of combined anti- γ interferon antibody and cyclosporine therapy on cardiac allograft survival in the rat. *Transplantation* 1988; 45:222-223
97. Iwai H, Kuma SI, Inaba MM, et al: Acceptance of murine thyroid allografts by pretreatment of anti-Ia antibody or anti-dendritic cell antibody in vitro. *Transplantation* 1989; 47:45-49
98. Norman DJ: The clinical role of OKT3. *Immunol Allergy Clin North Am* 1989; 9:95-107
99. Weimar W, Baumgartner D, Hendriks GF, et al: The prophylactic use of Orthoclone OKT3 in kidney and heart transplantation. *Transplant Proc* 1988; 20(suppl 6):96-100

100. Debuire A, Chkoff N, Chatenoud L, et al: One-month prophylactic use of OKT3 in cadaver kidney transplant recipients. *Transplantation* 1988; 45:546-553
101. Stratta RJ, D'Alessandro AM, Hoffmann RM, et al: Cadaveric renal transplantation in the cyclosporine and OKT3 eras. *Surgery* 1988; 104:606-615
102. Norman DJ, Shield CF, Barry JM, et al: Therapeutic use of OKT3 monoclonal antibody for acute renal allograft rejection. *Nephron* 1987; 46(suppl 1):41-47
103. Fung JJ, Demetris AJ, Porter KA, et al: Use of OKT3 with cyclosporin and steroids for reversal of acute kidney and liver allograft rejection. *Nephron* 1987; 46(suppl 1):19-39
104. Ortho Multicenter Transplant Study Group: A randomized clinical trial of OKT3 monoclonal antibody for acute rejection of cadaveric renal transplants. *N Engl J Med* 1985; 313:337-342
105. Goldstein G: Monoclonal antibody specificity: Orthoclone OKT3 T-cell blocker. *Nephron* 1987; 46(suppl 1):5-11
106. Kahana L, Baxter J: OKT3 rescue in refractory renal rejection. *Nephron* 1987; 46(suppl 1):34-40
107. Norman DJ, Barry JM, Bennett WM, et al: The use of OKT3 in cadaveric renal transplantation for rejection that is unresponsive to conventional anti-rejection therapy. *Am J Kidney Dis* 1988; 11:90-93
108. Rubin MF, Nghiem DD, Stachura I: Late steroid-resistant rejection response to OKT3. *Transplantation* 1988; 45:818-819
109. Delaney VB, Campbell WG, Nasr SA, et al: Efficacy of OKT3 monoclonal antibody therapy in steroid-resistant, predominantly vascular acute rejection. *Transplantation* 1988; 45:743-748
110. Cosimi AB: OKT3: First-dose safety and success. *Nephron* 1987; 46(suppl 1):12-18
111. Abramowicz D, Schandene L, Goldman M, et al: Release of tumor necrosis factor, interleukin-2 and gamma interferon in serum after injection of OKT3 monoclonal antibody in kidney transplant recipients. *Transplantation* 1989; 47:606-608
112. Debets JM, Leunissen KM, Van Hooff HJ, et al: Evidence of involvement of tumor necrosis factor in adverse reactions during treatment of kidney allograft rejection with antithymocyte globulin. *Transplantation* 1989; 47:487-492
113. Oh CS, Stratta RF, Fox BC, et al: Increased infections associated with the use of OKT3 for treatment of steroid-resistant rejection in renal transplantation. *Transplantation* 1988; 45:68-73
114. Chou S, Norman DJ: Effect of OKT3 antibody therapy on cytomegalovirus reactivation in renal transplant recipients. *Transplant Proc* 1985; 17:2755-2756
115. Lazarovits AI, Shield CF: Recurrence of acute rejection in the absence of CD3-positive lymphocytes. *Clin Immunol Immunopathol* 1988; 48:392-400
116. Shield CF III, Norman DJ, Marlett P, et al: Comparison of antimouse and antihorse antibody production during the treatment of allograft rejection with OKT3 or antithymocyte globulin. *Nephron* 1987; 46(suppl 1):48-51
117. Chatenoud L, Jonker M, Villemain F, et al: The human immune response to the OKT3 monoclonal antibody is oligoclonal. *Science* 1986; 232:1406-1408
118. Norman DJ, Shield CF III, Henell KR, et al: Effectiveness of a second course of OKT3 monoclonal anti-T cell antibody for treatment of renal allograft rejection. *Transplantation* 1988; 46:523-529
119. First MR, Schroeder TJ, Hurtubise PE, et al: Successful retreatment of allograft rejection with OKT3. *Transplantation* 1989; 47:88-91
120. Mayes JT, Thistlethwaite JR, Stuart JK, et al: Reexposure to OKT3 in renal allograft recipients. *Transplantation* 1988; 45:349-353
121. Shield CF, Hughes JD, Marlett P, et al: The human anti-mouse response to OKT3—Crossreactive pattern analysis using a large group of anti-CD3 and isotypic monoclonal antibodies. *Transplant Proc* 1989; 21:981-984
122. Hirsch R, Chatenoud L, Gress RE, et al: Suppression of the humoral response to anti-CD3 monoclonal antibody. *Transplantation* 1989; 47:853-857
123. Paetkau V, Mills GB: Cytokines and mechanisms of lymphocyte activation. *Immunol Allergy Clin North Am* 1989; 9:21-43
124. Strom TB, Kelley VE: Toward more selective therapies to block undesired immune responses. *Kidney Int* 1989; 35:1026-1033
125. Diamantstein T, Osawa H, Kirkman RL, et al: Interleukin 2 receptor—A target for immunosuppressive therapy. *Transplant Rev (Orlando)* 1987; 1:177-196
126. Souillou JP, Peyronnet P, Le Mauff B, et al: Prevention of rejection of kidney transplants by monoclonal antibody directed against interleukin 2. *Lancet* 1987; 1:1339-1342
127. Kupiec-Weglinski JW, Diamantstein T, Tilney NL: Interleukin 2 receptor-targeted therapy—Rationale and applications in organ transplantation. *Transplantation* 1988; 46:785-792
128. Cantarovich D, Le Mauff B, Hourmant M, et al: Anti-interleukin 2 receptor monoclonal antibody in the treatment of ongoing acute rejection episodes in human kidney graft—A pilot study. *Transplantation* 1989; 47:454-457
129. Turka LA, Carpenter CB, Yunis EJ, et al: Selective sparing of suppressor cells generated in mixed lymphocyte response by an anti-interleukin-2 receptor antibody. *Transplantation* 1989; 47:182-188
130. Murphy JR, Kelley VE, Strom TB: Interleukin 2 toxin: A step toward selective immunomodulation. *Am J Kidney Dis* 1988; 11:159-162
131. Pankewycz O, Mackie J, Hassarjian R, et al: Interleukin-2-diphtheria toxin fusion protein prolongs murine islet cell engraftment. *Transplantation* 1989; 47:318-322
132. Kirkman RL, Bacha P, Barrett LV, et al: Prolongation of cardiac allograft survival in murine recipients treated with a diphtheria toxin-related interleukin-2 fusion protein. *Transplantation* 1989; 47:327-330
133. Tejani A, Lancman I, Pomrantz A, et al: Nephrotoxicity of cyclosporine A and cyclosporine G in a rat model. *Transplantation* 1988; 45:184-187
134. Yoshimura N, Matsui S, Hamashima T, et al: Effect of a new immunosuppressive agent, FK506, on human lymphocyte responses in vitro—I. Inhibition of expression of alloantigen-activated suppressor cells, as well as induction of alloreactivity. *Transplantation* 1989; 47:351-356
135. Yoshimura N, Matsui S, Hamashima T, et al: Effect of a new immunosuppressive agent, FK506, on human lymphocyte responses in vitro—II. Inhibition of the production of IL-2 and γ -IFN, but not B cell-stimulating factor 2. *Transplantation* 1989; 47:356-359
136. Warty V, Diven W, Cadoff E, et al: Brief communications: FK506: A novel immunosuppressive agent—Characteristics of binding and uptake by human lymphocytes. *Transplantation* 1988; 46:453-455
137. Thomson W: FK-506—How much potential? *Immunol Today* 1989; 10:6-9
138. Tuch BE, Lissing JE, Suranyi MG: Immunomodulation of human fetal cells by the fungal metabolite gliotoxin. *Immunol Cell Biol* 1988; 66:307-312
139. Oesterwitz H, Scholz D, Kaden J, et al: Photochemical donor pretreatment in clinical kidney transplantation—Preliminary report. *Urol Res* 1987; 15:211-213
140. Pollack MS, Short HD III, Young JB, et al: Brief communications: Graft stability in a heart transplant recipient whose immunosuppressive therapy was discontinued for 8 months. *Transplantation* 1988; 45:242-243
141. Hall BM: Tolerance and specific unresponsiveness in organ transplantation. *Immunol Allergy Clin North Am* 1989; 9:61-77
142. Barber WH, Diethelm A, Laskow DA, et al: Use of cryopreserved donor bone marrow in cadaver kidney allograft recipients. *Transplantation* 1989; 47:66-71
143. Myburgh JA, Meyers AM, Thomson PD, et al: Total lymphoid irradiation—Current status. *Transplant Proc* 1989; 21:826-828
144. Chow D, Saper V, Strober S: Renal transplant patients treated with total lymphoid irradiation show specific unresponsiveness to donor antigens in the mixed leukocyte reaction (MLR). *J Immunol* 1987; 138:3746-3750
145. Strober S, Dhillon M, Schubert M, et al: Acquired immune tolerance to cadaveric renal allografts. *N Engl J Med* 1989; 321:28-33
146. Saper V, Chow D, Engleman ED, et al: Clinical and immunological studies of cadaveric renal transplant recipients given total-lymphoid irradiation and maintained on low-dose prednisone. *Transplantation* 1988; 45:540-546
147. Pahl MV, Barton CH, Ulich T, et al: Nephrology consultant: Renal dysfunction and macroangiopathic changes in a renal transplant patient. *Am J Nephrol* 1988; 8:72-79
148. Hall BM: Cellular infiltrates in allografts. *Transplant Proc* 1987; 19:50-56
149. Waltzer WC, Miller F, Arnold A, et al: Value of percutaneous core needle biopsy in the differential diagnosis of renal transplant dysfunction. *J Urol* 1987; 137:1117-1121
150. Bishop GA, Hall BM, Duggin GG, et al: Immunopathology of renal allograft rejection analyzed with monoclonal antibodies to mononuclear markers. *Kidney Int* 1986; 29:708-717
151. Colvin RB: Diagnostic use in transplantation: Clinical applications of monoclonal antibodies in renal allograft biopsies. *Am J Kidney Dis* 1988; 11:126-130
152. Hall BM, Bishop GA, Duggin GG, et al: Increased expression of HLA-DR antigens on renal tubular cells in renal transplants: Relevance to the rejection response. *Lancet* 1984; 2:247-251
153. Häyry P, von Willebrand E, Ahonen J, et al: Aspiration cytology in organ transplantation. *Transplant Rev (Orlando)* 1987; 1:133-157
154. Bishop GA, Hall BM, Waugh J, et al: Diagnosis of renal allograft rejection by analysis of fine-needle aspiration biopsy specimens with immunostains and simple cytology. *Lancet* 1986; 2:645-650
155. Tötterman TH, Hanäs E, Bergström R, et al: Immunologic diagnosis of kidney rejection using FACS analysis of graft-infiltrating functional and activated T and NK cell subsets. *Transplantation* 1989; 47:817-823
156. Dunbar KR, Salomon DR, Kaude J, et al: Loss of corticomedullary demarcation on magnetic resonance imaging: An index of biopsy-proven acute renal transplant dysfunction. *Am J Kidney Dis* 1988; 12:200-207
157. Van Gansbake D, Segebarth C, Toussaint C, et al: Non-obstructive kidney transplant dysfunction: Magnetic resonance evaluation. *Br J Radiol* 1988; 61:473-491
158. Te Straké L, Schultze Kool LJ, Paul LC, et al: Magnetic resonance imaging of renal transplants: Its value in the differentiation of acute rejection and cyclosporin A nephrotoxicity. *Clin Radiol* 1988; 38:220-228
159. Dubovsky FV, Russell CDJ: Radionuclide evaluation of renal transplants. *Semin Nucl Med* 1988; 18:181-198
160. Goldsmith MS, Tanasescu DE, Waxman AD, et al: Comparison of magnetic resonance imaging and radionuclide imaging in the evaluation of renal transplant failure. *Clin Nucl Med* 1988; 13:250-257
161. Hall JT, Kim EE, Pjura GA, et al: Correlation of radionuclide and ultrasound studies with biopsy findings for the diagnosis of renal transplant rejection. *Urology* 1988; 32:172-179
162. Buckley AR, Cooperberg PL, Reeve CE, et al: The distinction between acute renal transplant rejection and cyclosporine nephrotoxicity: Value of duplex sonography. *AJR* 1987; 149:521-525
163. Gankins SM, Sanfilippo FP, Carroll RA: Duplex doppler sonography of renal transplants: Lack of sensitivity and specificity in established pathologic diagnosis. *AJR* 1989; 152:535-539
164. Allen KS, Jorkasky DK, Arger PH, et al: Renal allografts: Prospective analysis of Doppler sonography. *Radiology* 1988; 169:371-376
165. Foxwell BMJ, Ryffel B: The mechanisms of action of cyclosporine. *Immunol Allergy Clin North Am* 1989; 9:79-93
166. Sommer BG, Sing DE, Henry ML, et al: Serum cyclosporine kinetic profile—Failure to correlate with nephrotoxicity or rejection episodes following sequential immunotherapy for renal transplantation. *Transplantation* 1988; 45:86-90
167. Mihatsch MJ, Steiner K, Abeywickrama KH, et al: Risk factors for the development of chronic cyclosporine nephrotoxicity. *Clin Nephrol* 1988; 29:165-175
168. Moyer TP, Post GR, Sterioff S, et al: Cyclosporine nephrotoxicity is minimized by adjusting dosage on the basis of drug concentration in blood. *Mayo Clin Proc* 1988; 63:241-247
169. Lorber MI, Flechner SM, Van Buren CT, et al: Cyclosporine toxicity: The

effect of combined therapy using cyclosporine, azathioprine, and prednisone. *Am J Kidney Dis* 1987; 6:476-484

170. Rocher LL, Hodgson RJ, Merion RM, et al: Amelioration of chronic renal allograft dysfunction in cyclosporine-treated patients by addition of azathioprine. *Transplantation* 1989; 47:249-254

171. Nahman DS, Cosio FG, Kolkin S, et al: Cyclosporine nephrotoxicity without major organ transplantation. *Ann Intern Med* 1987; 106:400-402

172. Ontario Renal Transplant Research Group: Factors influencing early renal function in cadaveric kidney transplants—A case control study. *Transplantation* 1988; 45:122-127

173. Canadian Multicenter Transplant Study Group: A randomized clinical trial of cyclosporine in cadaveric renal transplantation. *N Engl J Med* 1983; 309:809-815

174. Halloran PF, Aprile MW, Farewell V, et al: Early function as the principal correlate of graft survival—A multivariate analysis of 200 cadaveric renal transplants treated with a protocol incorporating antilymphocyte globulin and cyclosporine. *Transplantation* 1988; 46:223-228

175. Sommer BG, Henry MC, Ferguson RM: Sequential antilymphoblast globulin and cyclosporine for renal transplantation. *Transplantation* 1987; 43:85-90

176. Golub MS, Berger ME: Direct augmentation by cyclosporine A of the vascular contractile response to nerve stimulation. *Hypertension* 1987; 9(pt 2):III:96-100

177. Curtis JJ, Luke RG, Dubovsky E, et al: Cyclosporin in therapeutic doses increases renal allograft vascular resistance. *Lancet* 1986; 2:477-479

178. English J, Evan A, Houghton DC, et al: Cyclosporine-induced renal dysfunction in the rat: Evidence for arteriolar vasoconstriction with preservation of tubular function. *Transplantation* 1987; 44:135-141

179. Remuzzi G, Bertani T: In-depth review: Renal vascular and thrombotic effects of cyclosporine. *Am J Kidney Dis* 1989; 13:261-272

180. Bennett WM: Basic mechanisms and pathophysiology of cyclosporine nephrotoxicity. *Transplant Proc* 1985; 17:297-302

181. Rogers TS, Elzinga L, Bennett WM, et al: Selective enhancement of thromboxane in macrophages and kidneys in cyclosporine-induced nephrotoxicity—Dietary protection by fish oil. *Transplantation* 1988; 45:153-156

182. Petric R, Freeman D, Wallace C, et al: Effect of cyclosporine on urinary prostanoid excretion, renal blood flow and glomerulotubular function. *Transplantation* 1988; 45:883-889

183. Voss BL, Hamilton KK, Samara ENS, et al: Cyclosporine suppression of endothelial prostacyclin generation. *Transplantation* 1988; 45:793-796

184. Coffman TM, Carr DR, Yarger WE, et al: Evidence that renal prostaglandin and thromboxane production is stimulated in chronic cyclosporine nephrotoxicity. *Transplantation* 1987; 43:282-285

185. Perico N, Benigni A, Zoja C, et al: Functional significance of exaggerated renal thromboxane A₂ synthesis induced by cyclosporin A. *Am J Physiol* 1986; 251 (pt 2):F581-587

186. Elzinga L, Kelley V, Houghton DC, et al: Fish oil modifies experimental cyclosporine nephrotoxicity and decreases renal prostaglandins. *Transplantation* 1987; 43:271-273

187. Baciewicz AM, Baciewicz FA: Cyclosporine pharmacokinetic drug interactions. *Am J Surg* 1989; 157:264-271

188. Barros EJG, Boim MA, Ajzen H, et al: Glomerular hemodynamics and hormonal participation on cyclosporine nephrotoxicity. *Kidney Int* 1987; 32:19-25

189. Handschumacher RE, Harding MW, Rice J, et al: Cyclophilin: A specific cytosolic binding protein for cyclosporin A. *Science* 1984; 226:544-547

190. Lewis RM, Janney RP, Golden DL, et al: Stability of renal allograft function associated with long-term cyclosporine immunosuppressive therapy—Five-year follow-up. *Transplantation* 1989; 47:266-272

191. Myers BD, Sibley R, Newton L, et al: The long-term course of cyclosporine-associated chronic nephropathy. *Kidney Int* 1988; 33:590-600

192. McDiarmid SV, Ettenger RB, Fine RN, et al: Serial decrease in glomerular filtration rate in long-term pediatric liver transplantation survivors treated with cyclosporine. *Transplantation* 1989; 47:314-318

193. Ruiz P, Kolbeck PC, Scroggs MW, et al: Associations between cyclosporine therapy and interstitial fibrosis in renal allograft biopsies. *Transplantation* 1988; 45:91-95

194. Canadian Multicentre Transplant Study Group: A randomized clinical trial of cyclosporine in cadaveric renal transplantation: Analysis at three years. *N Engl J Med* 1986; 314:1219-1225

195. Tomlanovich S, Golbetz H, Perlroth M, et al: Limitations of creatinine in quantifying the severity of cyclosporine-induced chronic nephropathy. *Am J Kidney Dis* 1986; 8:332-337

196. Curtis JJ: Hypertension in the renal transplant patient. *Transplant Rev (Orlando)* 1988; 2:17-28

197. Luke RG: Nephrology forum: Hypertension in renal transplant recipients. *Kidney Int* 1987; 31:1024-1037

198. Chapman JR, Marcen R, Arias M, et al: Hypertension after renal transplantation—A comparison of cyclosporine and conventional immunosuppression. *Transplantation* 1987; 43:860-864

199. Rosansky SJ, Sugimoto T: An analysis of the United States renal transplant patient population and organ survival characteristics: 1977 to 1980. *Kidney Int* 1982; 22:685-692

200. McGeown MG, Douglas JF, Donaldson RA, et al: Ten-year results of renal transplantation with azathioprine and prednisolone as only immunosuppression. *Lancet* 1988; 1:983-985

201. Cheigh JS, Haschemeyer RH, Wang JCL, et al: Hypertension in kidney transplant recipients—Effect on long-term renal allograft survival. *Am J Hypertens* 1989; 2(pt 1):341-348

202. Curtis JJ, Luke RG, Diethelm AG, et al: Benefits of removal of native kidneys in hypertension after renal transplantation. *Lancet* 1985; 2:739-742

203. Linas SL, Miller PD, McDonald KM, et al: Role of the renin-angiotensin

system in post-transplantation hypertension in patients with multiple kidneys. *N Engl J Med* 1978; 298:1440-1444

204. Thompson ME, Shapiro AP, Johnsen AM, et al: The contrasting effects of cyclosporin-A and azathioprine on arterial blood pressure and renal function following cardiac transplantation. *Int J Cardiol* 1986; 11:219-229

205. Palestine AG, Nussenblatt RB, Chan CC: Side effects of systemic cyclosporine in patients not undergoing transplantation. *Am J Med* 1984; 77:652-656

206. Myers BD: Cyclosporine nephrotoxicity. *Kidney Int* 1986; 30:964-974

207. Von Graffenried B, Krupp P: Side effects of cyclosporine in renal transplant recipients and in patients with autoimmune diseases. *Transplant Proc* 1986; 18:876-883

208. Curtis JJ, Luke RG, Jones P, et al: Hypertension in cyclosporine-treated renal transplant recipients is sodium dependent. *Am J Med* 1988; 85:134-138

209. Forman SJ, Textor SC, Carlson JE: Prednisone potentiates cyclosporine-induced blood pressure changes in normotensive bone marrow transplant recipients (Abstr). *Kidney Int* 1987; 31:297

210. Mahony JF, Sheil AGR: Long-term complications of cadaver renal transplantation. *Transplant Rev (Orlando)* 1987; 1:47-58

211. Ho M, Jaffe R, Miller G, et al: The frequency of Epstein-Barr virus infection and associated lymphoproliferative syndrome after transplantation and its manifestations in children. *Transplantation* 1988; 45:719-727

212. Preiksaitis JK: Cytomegalovirus infection in transplant recipients. *Immunol Allergy Clin North Am* 1989; 9:137-151

213. Lewis RM, Johnson PC, Golden D, et al: The adverse impact of cytomegalovirus infection on clinical outcome in cyclosporine-prednisone treated renal allograft recipients. *Transplantation* 1988; 45:353-359

214. Boyce NW, Hayes K, Gee D, et al: Cytomegalovirus infection complicating renal transplantation and its relationship to acute transplant glomerulopathy. *Transplantation* 1988; 45:706-709

215. Grundy JE, Lui SF, Super M, et al: Symptomatic cytomegalovirus infection in seropositive kidney recipients: Reinfection with donor virus rather than reactivation of recipient virus. *Lancet* 1988; 2:132-135

216. Weir MR, Henry ML, Blackmore M, et al: Incidence and morbidity of cytomegalovirus disease associated with a seronegative recipient receiving seropositive donor-specific transfusion and living-related donor transplantation. *Transplantation* 1988; 45:111-116

217. Johnson PC, Lewis RM, Golden DL, et al: The impact of cytomegalovirus infection on seronegative recipients of seropositive donor kidneys versus seropositive recipients treated with cyclosporine-prednisone immunosuppression. *Transplantation* 1988; 45:116-121

218. Metselaar HJ, Ploeg RJ, Van Loon AM, et al: Prevention of CMV infection by screening for CMV antibodies in renal allograft recipients and the blood and kidney donors. *Scand J Infect Dis* 1988; 20:135-139

219. Balfour HH, Welo PK, Sachs GW: Cytomegalovirus vaccine trial in 400 renal transplant candidates. *Transplant Proc* 1985; 17:81-83

220. Farrar GH, Bull JR, Greenaway PJ: Prospects for the clinical management of human cytomegalovirus infections. *Vaccine* 1986; 4:217-224

221. Brayman KL, Dafoe DC, Smythe WR, et al: Prophylaxis of serious cytomegalovirus infection in renal transplant candidates using live human cytomegalovirus vaccine—Interim results of a randomized trial. *Arch Surg* 1988; 123:1502-1508

222. Sydnman DR, Werner BG, Tilney NL, et al: A further analysis of primary cytomegalovirus disease prevention in renal transplant recipients with a cytomegalovirus immune globulin: Interim comparison of a randomized and an open-label trial. *Transplant Proc* 1988; 20(suppl 8):24-30

223. Balfour HH, Chace BA, Stapleton JT, et al: A randomized placebo-controlled trial of oral acyclovir for the prevention of cytomegalovirus disease in recipients of renal allografts. *N Engl J Med* 1989; 320:1381-1387

224. Van der Bij W, Schirm J, Torensma R, et al: Comparison between viremia and antigenemia for detection of cytomegalovirus in blood. *J Clin Microbiol* 1988; 26:2531-2535

225. Kemnitz J, Haverich A, Gubernatis G, et al: Rapid identification of viral infections in liver, heart, and kidney allograft biopsies by in situ hybridization (Letter). *Am J Surg Pathol* 1989; 13:80-82

226. DeBure A, Celton JL, Cartron J, et al: Granulocyte-associated immunoglobulins in renal transplant recipients with cytomegalovirus infection. *Lancet* 1988; 2:1338-1340

227. Verheyden JP: Evolution of therapy for cytomegalovirus infection. *Rev Infect Dis* 1988; 10(suppl 3):477-489

228. Cantarovich M, Hiesse C, Lantz O, et al: Treatment of cytomegalovirus infections in renal transplant recipients with 9-(1,3-dihydroxy-2-propoxymethyl)guanine. *Transplantation* 1988; 45:1139-1141

229. Stoffel M, Gianello P, Squifflet JP, et al: Effect of 9-(2-hydroxy-1-[hydroxymethyl] ethoxymethyl) guanine (DHPG) on cytomegalovirus pneumonitis after renal transplantation. *Transplantation* 1988; 46:594-595

230. Sydnman DR: Ganciclovir therapy for cytomegalovirus disease associated with renal transplants. *Rev Infect Dis* 1988; 10(suppl 3):554-562

231. Ringden O, Lönnqvist B, Paulin T, et al: Pharmacokinetics, safety and preliminary clinical experiences using foscarnet in the treatment of cytomegalovirus infections in bone marrow and renal transplant recipients. *J Antimicrob Chemother* 1986; 17:373-387

232. Aulitzky WE, Tilg H, Niederwieser D, et al: Ganciclovir and hyperimmunoglobulin for treating cytomegalovirus infection in bone marrow transplant recipients (Letter). *J Infect Dis* 1988; 158:488-489

233. Goodman JL: Possible transmission of herpes simplex virus by organ transplantation. *Transplantation* 1989; 47:609-613

234. Griffin PJA, Colbert JW, Williamson EPM, et al: Oral acyclovir prophylaxis of herpes infections in renal transplant recipients. *Transplant Proc* 1985; 17:84-85

235. Rubin RH, Jenkins RL, Shaw BW, et al: The acquired immunodeficiency syndrome and transplantation. *Transplantation* 1987; 44:1-4

236. Dummer JS, Erb S, Breinig MK, et al: Infection with human immunodeficiency virus in the Pittsburgh transplant population—A study of 583 donors and 1043 recipients, 1981-1986. *Transplantation* 1989; 47:134-139
237. Kumar P, Pearson JE, Martin DH, et al: Transmission of human immunodeficiency virus by transplantation of a renal allograft, with development of the acquired immunodeficiency syndrome. *Ann Intern Med* 1987; 106:244-245
238. Carbone LG, Cohen DJ, Hardy MA, et al: Determination of acquired immunodeficiency syndrome (AIDS) after renal transplantation. *Am J Kidney Dis* 1988; 11:387-392
239. Rubin H, Tolkoff-Rubin NE: The problem of human immunodeficiency virus (HIV) infection and transplantation. *Transplant Int* 1988; 1:36-42
240. Schwarz A, Hoffmann F, L'age-Stehr J, et al: Human immunodeficiency virus transmission by organ donation—Outcome in cornea and kidney recipients. *Transplantation* 1987; 44:21-24
241. Oliveira DB, Winearls CG, Cohen J, et al: Brief communications: Severe immunosuppression in a renal transplant recipient with HTLV-III antibodies. *Transplantation* 1986; 41:260-263
242. Sheil AG: Cancer after transplantation. *World J Surg* 1986; 10:389-396
243. Penn I: Transmission of cancer with donor organs. *Transplant Proc* 1988; 20:739-740
244. Sheil AGR, Flavel S, Disney APS, et al: Cancer development in patients progressing to dialysis and renal transplantation. *Transplant Proc* 1985; 17:1685-1689
245. Sheil AGR, Flavel S, Disney APS, et al: Cancer incidence in renal transplant patients treated with azathioprine or cyclosporin. *Transplant Proc* 1987; 19:2214-2216
246. McLelland J, Chu AC: Skin tumours in renal allograft recipients. *J R Soc Med* 1989; 92:110-111
247. Hanto DW, Frizzera G, Gajl-Peczalska KJ, et al: Epstein-Barr virus, immunodeficiency and B-cell lymphoproliferation. *Transplantation* 1985; 39:461-472
248. Ho M, Miller G, Atchison RW, et al: Epstein-Barr virus infections and DNA hybridization studies in posttransplantation lymphoma and lymphoproliferative lesions: The role of primary infection. *J Infect Dis* 1985; 152:876-886
249. Nalesnik MA, Jaffe R, Starzl TE, et al: The pathology of posttransplant lymphoproliferative disorders occurring in the setting of cyclosporine A-prednisone immunosuppression. *Am J Pathol* 1988; 133:173-192
250. Toussaint C, Kinnaert P, Vereerstraeten P: Late mortality and morbidity five to eighteen years after kidney transplantation. *Transplantation* 1988; 45:554-558
251. Rao KV, Andersen RC: Long-term results and complications in renal transplant recipients—Observations in the second decade. *Transplantation* 1988; 45:45-52
252. Rao KV, Kasiske BL, Bloom PM: Acute graft rejection in the late survivors of renal transplantation—Clinical and histological observations in the second decade. *Transplantation* 1989; 47:290-292
253. Sibley RV, Snover DC: The use and interpretation of biopsies in the management of the post-transplant patient, chap 26, *In* Cerilli GJ, (Ed): *Organ Transplantation and Replacement*. Philadelphia, JB Lippincott, 1988, pp 394-422
254. Ogura S, Banner B, Zerbe T, et al: Participation of dendritic cells in vascular lesions of chronic rejection of human allografts. *Lancet* 1988; 2:933-936
255. Foster MC, Wanham PW, Rowe PA, et al: The late results of renal transplantation and the importance of chronic rejection as a cause of graft loss. *Ann R Coll Surg Engl* 1989; 71:44-71
256. Rovelli M, Palmeri D, Vossler E, et al: Noncompliance in organ transplant recipients. *Transplant Proc* 1989; 21(pt 1):833-841
257. Didlake RH, Dreyfus K, Kerman RH, et al: Patient noncompliance: A major cause of late graft failure in cyclosporine-treated renal transplants. *Transplant Proc* 1988; 20(suppl 3):63-69
258. Fine RN: The adolescent with end-stage renal disease. *Am J Kidney Dis* 1985; 6:81-85
259. Reznik VM, Jones KL, Durham BL, et al: Changes in facial appearance during cyclosporin treatment. *Lancet* 1987; 1:1405-1407
260. Stewart RS: Psychiatric issues in renal dialysis and transplantation. *Hosp Community Psychiatry* 1983; 34:623-681
261. Korbet SM, Schwartz MM, Lewis EJ: Recurrent nephrotic syndrome in renal allografts. *Am J Kidney Dis* 1988; 11:270-276
262. Berger J: Recurrence of IgA nephropathy in renal allografts. *Am J Kidney Dis* 1988; 12:371-372
263. Mathew TH: Recurrence of disease following renal transplantation. *Am J Kidney Dis* 1988; 12:85-96
264. Brensilver JM, Mallat S, Scholes J, et al: Recurrent IgA nephropathy in living-related donor transplantation: Recurrence or transmission of a familial disease? *Am J Kidney Dis* 1988; 12:147-151
265. Vincenti F, Biava C, Tomlanovitch S, et al: Inability of cyclosporine to completely prevent the recurrence of focal glomerulosclerosis after kidney transplantation. *Transplantation* 1989; 47:595-598